# ALLELOPATHIC POTENTIAL OF PLANT RESIDUES OF THIRTY SRI LANKAN TRA-DITIONAL RICE (*Oryza sativa* L.) VARIETIES ON BARNYARDGRASS (*Echinochloa crus-galli*)

DL Wathugala\* and AS Ranagalage

Department of Crop Science, Faculty of Agriculture, University of Ruhuna, Mapalana, Kamburupitiya, Sri Lanka

Accepted: 05th February 2015

**ABSTRACT** 

The use of rice (Oryza sativa L.) allelopathy for weed management could reduce the need for applying herbicides to the crop. In this study a pot experiment and a laboratory bioassay were conducted to assess the allelopathic potential of residues of 30 traditional rice varieties in Sri Lanka on seed germination and seedling growth of barnyardgrass (Echinochloa crus-galli). All seeds were collected from "Farmer Federation for Conservation of Traditional Seeds and Agri-Resources". Rice residues were incorporated with silica sand (1% w/w) in small pots to prepare residue mixture and ground rice residues mixed with water (2.5g/100ml) were used to prepare residue extract for laboratory bioassay. Rice varieties exhibited marked differences in the inhibition of barnyardgrass growth and development. In the bioassay, residue extract of rice variety Herathbanda exhibited the greatest activity on the weed seedling growth, reducing dry weight by 30.7%, while Gonabaru residue extract exhibited the greatest inhibition on seed germination, reducing germination by 29.6%. In the pot experiment of residue mixture, Batapola el and Sula varieties showed the greatest inhibition percentage on germination (48.4%). However, Godaheenati showed the greatest inhibition % on plant height (27.4%) and dry weight (45.6%). These results suggest that incorporation of rice residues of tested varieties has influence on controlling barnyardgrass seedling growth.

Key words: Allelopathy, Oryza sativa L., Echinochloa crus-gall, traditional rice, rice residue

### INTRODUCTION

Allelopathy is defined as the direct or indirect harmful or beneficial effects of one plant on another through the production of chemical compounds that escape into the environment (Rice, 1984). Chemicals released from plants and imposing allelopathic influences are termed allelochemicals. Allelochemicals can be present in several parts of plants including roots, rhizomes, leaves, stems, pollen, seeds and flowers. Allelochemicals are released into the environment by root exudation, leaching from aboveground parts and volatilization and/ or by decomposition of plant material (Rice, 1984; Reigosa *et al.*, 1999). This phenomenon could be an alternative weed control method.

Rice (Oryza sativa L.) is the staple food of Sri

Lankans and one of the most widely grown food crops in Sri Lanka. In all rice ecosystems, herbicides have become one of the most important components in weed control due to the adoption of high yielding varieties and availability of cheap herbicides. However, intensive and repeated application of herbicides has resulted various environmental problems such as environmental pollution, evolving resistant weeds, (Valverde et al., 2000), residual effects on following crop and disappearance of susceptible weeds which effects weed biodiversity (Itoh, 2000). In this regard, an alternative to such a heavy dependence on herbicide is needed. Such an alternative might be found in the use of allelopathy, which can reduce herbicide use in rice cropping systems. Allelopathy can be used in weed control in two ways (1):

<sup>\*</sup>Corresponding author: lakmini077@yahoo.com

Selecting an appropriate crop varieties or incorporating allelopathic characters in to a desired crop variety, and (2): Applying residues or straw as mulches or growing an allelopathic varieties in a rotational sequence (Rice, 1995).

Barnyardgrass (*Echinochloa crus-galli*) is one of the greatest yield-limiting weeds in the irrigated rice systems of Sri Lanka. It is better adapted to grow under dry conditions rather than wet conditions. Now most paddy farmers practice direct seeding to minimize cost of production and due to water scarcity paddy fields are not frequently in flooded condition. These practices create good conditions to grow barnyardgrass in the paddy field and this weed expect to become greater problem in future (Im *et al.*, 1993).

Therefore, the most environmentally acceptable and sustainable approach is utilization of allelopathy for control of barnyardgrass and other paddy weeds. A number of studies have been conducted to evaluate the allelopahic potential from rice germplasm and number of rice varieties having allopathic potential have been determined in different places (Ahn and Chung, 2000; Chou, 1995, 1999; Chung et al., 1997; Dilday et al., 1994; Fujii, 1992; Garrity et al., 1992; Hassn et al., 1994; Lin et al., 1992; Olofsdotter et al., 1995). As an example Dilday et al. (1998) identified 412 varieties having allelopathic potential against ducksalad (Heteranthera limosa (Sw.) Willd.), among 12,000 varieties that originated from 31 different countries. These studies showed allelopathic potential is widely present in rice germplasm. Further it is assumed that rice allelopathy might be polygenetically controlled because it shows a continuous variation in the germplasm (Kim and Shin, 2003). Moreover, allelopathic potential is often attributed to several inhibitors that are assumed to act in an additive or synergistic way rather than in an isolated way (Courtois and Olofsdotter, 1998). Allelopathic potential of rice residues has also been studied in several places (Chung et al., 2001a; Chung et al., 2001b; Chung et al., 2002) and several allelochemicals have also been extracted from rice residues. Although allelopathic research has been conducted for several decades very limited knowledge is still available. Traditional rice varieties have been conserved, developed and used by Sri Lankan farmers over a period of more than 3000 years. However, with the introduction of high yielding varieties along with modern agronomic practices most traditional rice varieties were gradually replaced by small number of high yielding rice varieties. This was the main cause of narrow genetic diversity of cultivating rice. Although scientifically not proved different historical records reveal that traditional rice varieties have various nutritional and medicinal properties. Therefore, now there is a high demand for traditional rice and because of that there is a trend in cultivating traditional varieties by small scale farmers. Plant Genetic Resource Center (PGRC) Gannoruwa, the pioneer institute of genetic resource conservation in Sri Lanka has conserved more than 2000 traditional rice varieties of Sri Lanka. Apart from them some farmer federations such as 'Farmer Federation for Conservation of Traditional Seeds and Agri-Resources (FFCTS&AR)' are also cultivating and conserving traditional rice varieties with the help of PGRC. The main purpose of this study was to assess the allelopathic potential of residues of some traditional rice varieties grown by Sri Lankan farmers on barnyardgrass seed germination and seedling growth.

## MATERIALS AND METHODS

Seeds of traditional rice varieties were collected from "Farmer Federation for Conservation of Traditional Seeds and Agri-Resources", Homagama. Those cultivars were grown in pots at Faculty of Agriculture, University of Ruhuna and harvested in March 2014. The harvested plants were dried at room temperature (28°C). Leaves and stem parts were ground into a fine powder and it was used to prepare residue mixture for pot experiment and residue extract for bioassay.

**Pot experiment:** According to Jung *et al.* (2004)1% w/w, ground rice residues was used to prepare residue mixture for pot experiment. Three grams of residue was mixed thoroughly with 300 g of silica sand in each pot (diameter 9cm and height 10cm). Fifty barnyardgrass seeds were placed uniformly 1cm deep in each pot after 2 weeks of residue incorporation. The emergence percent was defined as the coleoptile protrusion through the soil surface and was counted each day for 7 days after planting. After emergence, seedlings were thinned to 20 plants per pot. Water was added to each pot to maintain adequate moisture. All plants were harvested 20 days after planting (DAP). All plants from each pot were measured for plant height. After the measurements were made, the seedlings were dried at 65°C for 8 h for dry weight determination. Control plants were grown in silica sand without residue. Average inhibition percentage was calculated getting average of inhibition percentages of seed germination (GP), Plant height (PH) and dry weight (DW). The inhibitory percent was calculated using the following equation.

Percentage Inhibition (%) = 
$$\frac{(C-R)}{C} \times 100$$

C:- Control value

R:- Residue mixture value

Experiment was conducted in a protected house with an average temperature of 28°C.

Bioassay: Aqueous extracts (w/v) were prepared by using the method described by Chung et al., (2001b). Two point five grams (2.5g) of finely ground rice residues mixed with 100 ml distilled water and shake (250rpm) for 1 h. Then the solutions were filtered through four layers of filter papers to remove fiber debris and stored in a refrigerator (4°C). Fifty barnyardgrass seeds were placed on a filter paper (Whatman No. 42) in a 4cm petri dish. Filter papers were wetted by adding similar volume of extract solutions. Distilled water was used as the control. The Petri dishes were placed in room temperature (≈28°C). Everyday similar amount of extracts were added to moist filter papers. Germination was determined after 7 days. Seedling root and shoot lengths were also measured after 7 days, then seedlings was oven dried at 65°C for 4 h for dry weight determination. Average inhibition percentage was calculated getting average of inhibition percentages of seed germination (GP), Shoot length (SL), Root length (RL) and dry weight (DW). The inhibitory percent was calculated using equation described under pot experiment.

Statistical analysis: The experiment was conducted two times with three replications using a randomized complete block design. The analysis of variance for all data was measured using the general linear model procedure of

the statistical analysis system (SAS). The mean values were separated on the basis of least significant difference (LSD) at the 0.05 probability level.

# RESULTS AND DISCUSSION

Rice varieties used in this experiment showed wide variations in inhibition of barnyardgrass seed germination (GP), plant height (PH) seedling dry weight (DW) and average inhibition (AI) (table 1). Inhibition of GP was greater than that of other measured parameters in pot experiment (table 1 and figure 1). Whereas the inhibitory effect was lowest on barnyardgrass shoot length (figure 1, table 1). Rice varieties Hichchinangi and Duruwee are the most (53.1%) and least (1.3%) inhibition of barnyardgrass seed germination respectively (table 1). However, rice variety Hichchinangi showed stimulation on PH and DW accumulation, which caused average stimulatory effect on barnyardgrass growth. Rice variety *Pokkali* was the most (47.4%) inhibitory of barnyardgrass dry weight accumulation, whereas, lowest inhibition (1.5%) caused by Kaluheenati. Godaheenati and Gurusinghewee showed highest (27.4%) and lowest (2.8%) inhibition in plant height respectively (table 1). All rice varieties used for this experiment except Hichchinangi showed positive average inhibition on barnyardgrass growth. The average inhibition of barnyardgrass from residue mixture of 30 rice varieties was highest for Godaheenati (28.2%)and lowest for Gurusinghewee (0.7%).

All varieties used for bioassay showed inhibitory effect on seed germination of barnyard grass. In particular rice variety *Kuruluthuda* 

(28.8%) had highest germination inhibition and *Mudaliwee* (0.01%) had lowest inhibition. Greatest inhibition in dry matter accumulation was observed in rice variety *Hichchinangi* (38.9%) in bioassay experiment. *Herathbanda* also showed considerably higher (34.8%) inhibition in dry matter accumulation and it was not significantly differ from *Hichchinangi*. Highest root length inhibition in bioassay was observed in rice variety *Herathbanda* and variety *Dahanala* showed highest shoot length inhibition (table 1).

The inhibition of growth and development of barnyardgrass observed in this study may be resulted that toxic compounds released from the residues or produced by microorganism activity during residue decomposition. Residue aqueous extracts consists only water soluble alleochemicals. Potential allelochemicals has been extracted from soils incorporated with rice residue as well as residue aqueous extracts. As an example Chung et al. (2001a) has extracted 9 known allelochemicals and their mixtures from rice straw extracts by high performance liquid chromatography (HPLC) analysis. When susceptible plants are exposed to allelochemicals, germination, growth and development may be affected. The most frequent reported gross morphological effects on plants are retarded seed germination, effects on coleoptile elongation and on radicle, shoot and root development (Kruse, 2000). Weir et al. (2004) declared that inhibition of photosynthetic rate, interruption of respiration; ATP synthesis and amino acids metabolism were major physiological and biochemical mechanism that might be mediated by allelochemicals.

The magnitude of growth inhibition varied

among rice varieties studied and it may due to the differences in allelopathic characteristics of rice varieties. Chung et al. (1997), Olofsdotter et al. (1995), Dilday et al. (1994) and Ahn and Chung (2000) have conducted several experiments using rice residue extracts and rice residue incorporation with silica sand to compare allelopathic characteristics of various rice varieties and results showed that variations in allelopathic activity exist among tested rice varieties. A study conducted by Khan and Vaishya (1992) also reported the rice residue in soil inhibited the population and biomass of Echinochloa colona. The difference in response was attributed to the genetic differences among the varieties, since the amount of rice residue incorporated were same. However, this allelopathic activity may be a result of higher concentrations of same chemical or a combination of different chemicals. Because, there are possibilities in decomposing rice residues may produce different amount of one or more allelopathic substances.

Allelopathic potential would be a valuable trait to incorporate in rice cultivars for improved weed management. Among the rice varieties used in this study none of the varieties showed considerably higher inhibition percentage to use in breeding programmes. However, results revealed that incorporating residues with soil rice can reduce barnyardgrass growth in some extent and it is important to do further screenings using other traditional varieties and also with improved varieties to select rice varieties with higher allelopathic potential.

There is a belief that varieties with coloured pericarp have high nutraceutucal properties

than white rice. Therefore, it could also be assumed that rice varieties with coloured pericarp may have higher allelopathic properties. Therefore, an analysis was done to examine effect of pericarp colour on allelopathic potential of rice varieties used in this experiment and results showed in table 2. Jung et al., (2004) also showed that rice residues of varieties with coloured hull have higher allelopathic potential on barnyardgrass than varieties with colourless hull. However, contrasting results were observed in this experiment, ie. average inhibition of white rice is higher when residue mixed with sand but red rice showed higher performances in bioassay (table 2). Residue extract consist only water soluble allelochemicals and seedlings exposed considerably higher concentrations of these allelochemicals in bioassays than residue incorporate with sand. When residues incorporate with sand these water soluble chemicals release slowly and in low concentrations together with other chemicals release during decomposition. Therefore, red rice may consist considerably higher quantity of water soluble allelochemical and white rice tested in this experiment may release considerably greater amount of allelochemicals during decomposition process.

Inhibitory effects in terms of maturation time were examined and results showed in table 3. In residue mixture experiment 3½ months varieties exerted significantly higher percentages of average inhibition (19.0%) than 4 and 3 months old varieties (16.9% and 9.4% respectively). However, 4 months varieties showed significantly highest average inhibition on barnyardgrass growth in bioassay. In terms of seed germination inhibition percentage, 4 months varieties had significantly greatest in-

hibition percentage (37.2%) in residue mixture experiment, whereas 3 months varieties had significantly greatest inhibition in bioassay (table 3). These results indicate that the time of maturity may effect on allelopathic potential. However, most experimental evidences proved that allelopathic characters attribute with genetic differences of rice varieties. Several rice varieties used in this experiment showed stimulatory effects on shoot length, root length and dry weight but inhibitory effect on seed germination (var. Hichchinangi). In 1984, Rice reported that stimulatory effects can happen at low concentrations of allelopathic substances but inhibitory effects at higher concentrations. Out of 30 varieties tested in this experiment 7 varieties in bioassay and 15 varieties in residue incorporation experiment showed more than 20% average inhibition on barnyardgrass growth (figure 2) indicating the importance of residue incorporation to paddy soil. In this experiment 1% rice residue incorporated to prepare mixture. Therefore, 4688 kgha<sup>-1</sup> would be theoretically required for allelopathy to occur in the field. Moreover, other than amount of residue many factors are also involved in allelopathic activity such as soil conditions, climatic conditions, management conditions etc. Also, allelochemicals released from decomposing straw at field condition may not remain active long due to further microbial activity. Therefore, different results would be expected in field conditions. It can be said that the more rice residue remaining in the paddy soil, the greater the concentration of allelopathic substances released during decomposition and higher degree of weed control. However, further experiments with various amount of rice residues are important to examine does dependent behavior of allelopathic effect. Some farmers in Sri Lanka leave all residues in the field but some farmers still burn rice residues before field preparation for next season. Chou (1995, 1999) and Chung *et al.* (2001a), who identified several allelopathic chemicals such as p-hydroxybenzoic, ferulic, p-coumaricsyringic and salicylic acids from rice leaves and straws extracts, decomposing straw, and in rice soil.

Rice is the staple food of Sri Lankans and most farmers cultivate improved rice varieties and considerably low number of farmers cultivates traditional varieties. There is lack of information available on allelopathic potential of both improved and traditional varieties on paddy weeds in Sri Lanka. Therefore, the observations produced from this research would provide information to develop rice varieties with higher allelopathic activity. This study suggests that the allelopathic compounds released when decomposing rice residues of tested traditional varieties and can act as a natural herbicide to control weeds. Therefore, incorporation of rice residues to paddy soil can be used to reduce cost for weed control in direct seeding rice ecosystems. However, further careful experiments are needed to select allelopathic varieties under field conditions.

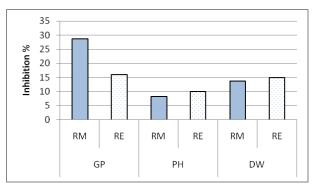


Figure 1: Inhibitory effect of rice residue mixture (RM) and residue extract (RE) on different parameters measured

Table 1: The inhibitory effects of 30 traditional rice varieties residue mixture and residue extract on barnyardgrass seed germination and seedling growth along with pericarp colour and duration taken to maturity\*

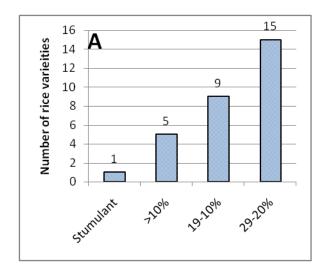
			Residue Mixture			Residue extract					
Variety name	PC	D(m)	GP	PH	DW	ΑI	GP	DW	RL	SL	ΑI
Murungakayan	R	4	27.3	8.2	13.6	16.4	17.6	23.5	19.7	14.3	18.8
Pachchaperum	R	3.5	35.6	12.2	19.9	22.5	5.6	4.2	-2.5	-7.4	0.0
Rathkaya	R	4	25.0	3.6	23.5	17.3	12.8	17.9	7.2	14.3	13.0
Behethheenati	R	3.5	40.2	-3.5	10.3	15.7	18.4	25.5	10.4	15.3	17.4
Sula	R	4	48.4	-6.9	-15.1	8.8	12.0	8.7	19.1	20.7	15.1
Galsyambala	R	3.5	36.7	10.1	8.1	18.3	21.6	19.1	5.8	11.7	14.5
Handiran	R	3.5	39.1	15.2	15.8	23.3	24.8	14.7	-4.8	-15.3	4.8
Batapola el	R	6	48.4	14.0	7.7	23.4	24.8	30.1	13.6	12.2	20.2
Rathkada	R	3	29.7	-1.9	-15.0	4.3	17.6	-38.9	-2.6	-35.1	-14.7
Madathawalu	R	4	40.6	24.0	-3.3	20.4	18.4	28.5	21.9	51.0	29.9
Gurusinghe wee	R	3.5	41.4	2.8	-42.2	0.7	18.4	6.6	-12.4	17.9	7.6
Gonabaru	R	3.5	20.3	4.2	18.3	14.3	29.6	28.8	2.1	16.8	19.3
Masuran	R	4.5	27.3	-4.1	-3.7	6.5	6.4	21.6	4.4	24.8	14.3
Herathbanda	R	3.5	39.1	-8.2	-6.3	8.2	18.4	34.8	38.1	31.6	30.7
Kaluheenati	R	4	39.1	3.6	1.5	14.7	24.0	33.2	9.0	-7.4	14.7
HichchiNangi	R	3	53.1	-15.2	-66.7	-9.6	22.4	38.9	1.4	32.2	23.7
Pokkali	R	3.5	10.3	-4.5	47.4	17.7	23.2	-4.8	-3.2	51.0	16.5
Kuruluthuda	R	3	21.5	10.4	-1.9	10.0	28.8	21.1	-0.9	20.6	17.4
Munchalalgi	R	3.5	37.9	5.1	4.2	15.7	18.4	32.5	0.3	13.5	16.2
Dahanala	R	4	43.0	11.0	17.7	23.9	22.4	20.9	7.4	36.5	21.8
Sudumurunga	W	3.5	15.4	18.1	33.3	22.3	7.2	17.9	-26.8	-16.6	-4.6
Sudurusamba	W	3	11.5	8.2	41.8	20.5	13.6	8.2	-0.1	3.8	6.4
Unakola samba	W	3.5	21.8	9.4	40.2	23.8	10.4	9.7	-14.5	-15.4	-2.5
Rathal	R	4.5	12.8	15.5	38.1	22.2	12.8	-15.5	12.3	29.2	9.7
Suwal	W	3.5	5.1	20.3	37.2	20.9	9.6	-6.5	-4.5	-32.7	-8.5
Hodarawalu	R	3.5	10.3	22.3	41.9	24.8	1.6	21.2	17.2	41.9	20.5
Duru wee	R	3	1.3	25.3	39.6	22.1	13.6	8.4	-14.1	-29.8	-5.5
Rathheenati	R	3.5	36.7	2.7	23.0	20.8	13.6	11.3	-11.4	-11.4	0.5
Godaheenati	R	3.5	11.5	27.4	45.6	28.2	11.2	13.1	9.3	26.9	15.1
Mudaliwee	W	3.5	27.4	18.3	34.6	26.7	0.01	11.1	1.4	-16.6	-1.0
CV			7.4	5.5	1.6	4.0	11.5	3.5	5.8	5.4	3.7
LSD (0.05%)			0.6	0.4	0.2	0.3	0.8	0.4	0.5	0.6	0.3

<sup>\*</sup>Data presented in the table are inhibition percentages PC, pericarp colour, R, red pericarp, W, white pericarp, D(m), Duration in months, GP, germination percentage, SL, shoot length, RL,root length, DW, dry weight, AI, average inhibition

Table 2:Inhibitory effect of pericarp colour of rice varieties on barnyard grass\*

Pericarp	Residu	ie mixt	ure						
colour	GP	PH	DW	AI GP	DW	RL	SL	ΑI	
Red rice	30.9a	7.4b	9.9b	16.0b	17.5a	17.0a	16.3a	5.9a	13.7a
White rice	13.5b	14.0	38.1a	21.9a	14.7b	14.5b	8.6b	0.9b	8.2b
CV	6.4	3.8	2	2.4	5.9	2.2	3.7	12.1	9.2
LSD	3.2	0.9	1.1	1	0.4	0.1	0.3	0.8	0.5

<sup>\*</sup> Data presented in the table are inhibition percentages. GP, germination percentage, PH, Plant height, DW, dry weight, SL, shoot length, RL,root length, AI, average inhibition.



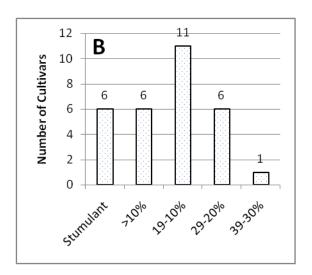


Figure 2: Distribution of rice varieties with allelopathic potential (based on average inhibition %) of rice residue mixture (A) and residue extract (B) on barnyardgrass seed germination and seedling growth.

Table 3: Inhibitory effect of duration to maturity of rice varieties on barnyardgrass\*

Maturation time	Residue	mixture			Residue (	Residue extract					
	GP	PH	DW	AI	GP	DW	SL	RL	ΑI		
4M	37.2a	7.2ab	6.3b	16.9b	17.9ab	22.1a	21.6a	14.0a	18.9a		
3.5 M	26.8b	9.5a	20.7a	19.0a	14.5b	14.9b	7.3b	-0.1b	9.1b		
3 M	23.4c	5.3b	-0.4c	9.4c	19.2a	7.5c	-1.6c	-3.2c	5.5b		
CV	1.9	7.9	5.1	2.1	5.3	2.9	18.2	18.1	7.2		
LSD (0.05%)	0.1	0.4	0.3	0.1	0.4	0.2	0.9	0.8	0.4		

<sup>\*</sup>Data presented in the table are inhibition percentages. GP, germination percentage, PH, Plant height, DW, dry weight, SL, shoot length, RL,root length, AI, average inhibition.

## **CONCLUSIONS**

Incorporating rice residue to paddy soil would be able to inhibit seed germination and growth of barnyardgrass to some extent. Results also revealed that the allelopathic characters differ among tested rice varieties. Further experiments with more traditional varieties and with improved rice varieties are important to select rice varieties with higher allelopathic potential. Isolation and identification of allelochemicals of selected varieties are also important for further analysis of allelopathic characters of rice. Because rice varieties with greater allelopathic potential might reduce application of herbicides by the farmers since these can lead us to identify varieties with a natural source of herbicides.

### **ACKNOWLEDGEMENTS**

The authors wish to acknowledge the financial support of the TURIS project throughout this experiment.

## REFERENCES

- Ahn JK and Chung IM 2000 Allelopathic potential of rice hulls on germination and seedling growth of barnyard grass. Agron. J. 92: 1162–1167.
- Chou CH 1995 Allelopathy and sustainable agriculture. In: Inderjit Dakshini KMM, Einhellig FA(Eds.), Allelopathy; Organisms, Process, and Application. ACS Symposium Series S82. American Chemical Society, Washington, DC, pp. 211–223.
- Chou CH 1999 Role of allelopathy in plant biodiversity and sustainable agriculture. Crit. Rev. Plant Sci. 18: 609–636.
- Chung IM, Kim KH, Ahn JK, Chun SC, Kim CS, Kim JT, Kim SH 2002 Screening of allelochemicals on barnyardgrass (*Echinochloa crus-galli*) and identification of potentially allelopathic compounds from rice (*Oryza sativa*) variety hull extracts. Crop Protection 21: 913–920.
- Chung IM, Ahn JK and Yun SJ 2001a Identification of allelopathic compounds from rice (*Oryza sativa L*.) straw and their biological activity. Can. J. Plant Sci. 81: 815–819.
- Chung IM, Ahn JK and Yun SJ 2001b Assessment of allelopathic potential of barnyard grass (*Echinochloa crusgalli*) on rice (*Oryza sativaL*.) cultivars Crop Protection 20: 921–928.
- Chung IM, Ahn JK, Kim JT, Kim CS, 2000 Assessment of allelopathic potentiality and identification of allelopathic compounds on Korean local rice varieties. Korean J. Crop Sci. 45:44–49.
- Chung IM, Kim KH, Ahn JK, Ju HJ 1997 Allelopathic potential evaluation of rice

- cultivars on *Echinochloa crus-galli*. Korean Weed Sci. 17: 52–58.
- Courtois B andOlofsdotter M1998 Incorporating the allelopathy trait in upland rice breeding programs.InOlofsdotter M eds. Allelopathy in Rice. Manila, Philippines: Int. Rice Research Institute: 57-68.
- Dilday RH, Yan WG, Moldenhauer KAK and Gravois KA1998Allelopathic activity in rice for controlling major aquatic weeds.InOlofsdotter M, eds.Allelopathy in Rice. Manila, Philippines: Int. Rice Research Institute: 7-26.
- Dilday RH, Lin J, Yan W 1994 Identification of allelopathy in the USDA-ARS rice germplasm collection. Aust. J. Exp. Agric. 34: 907–910.
- Fujii Y1992 The potential biological control of paddy weeds with allelopathyallelopathic effect of some rice cultivars. Proceedings of the International Symposium on Biological Control and Integrated Management of Paddy and Aquatic Weeds, Tsukuba, Japan: 305–320.
- Garrity DP, Movillon M, Moddy K1992 Differential weed suppression ability in upland rice cultivars. Agron. J. 84: 586 –591.
- Hassn SM, Rao AN, Bastawisi AO, Aidy IR1994 Weed management in broadcast seeded rice in Egypt. In: Moddy, Keds. Constraints, Opportunities and Innovations for Wet-Seeded Rice. IRRI Discussion Paper Series 10. Manila, Philippines: 257–269.
- Im IB, Guh OJ, Oh YJ1993 Weed occurrence and competitive characteristic under different cultivation types of rice

- (*OryzasativaL*.). Korean J. Weed Sci. 13: 114–121.
- Itoh K2000 Importance of biodiversity of aquatic plants in agro-ecosystem for rice production. In Abstracts of the Int. Symposium on Weed Biodiversity (National Sun Yat-sen University, Kaohsiung, Taiwan, 28-30 November, 2000). Weed Science Society of the Republic of China and National Sun Yat-sen University, Kaoshing:10.
- JungWS, KimKH, AhnJK, HahnSJ, ChungIM2004 Allelopathic potential of rice (*Oryza sativa L*.) residues against*Echinochloa crus-galli*. Crop Protection 23:211–218.
- Khan AH and Vaishya RD 1992 Allelopathic effects of different crop residues on germination and growth of weeds. In: proceedings of the First Nat. Sym. Allelopathy in agroecosystems (agriculture & forestry). Edited by P Tauro and SS Narwal. Indian society of Allelopathy. Haryana Agricultural University, Hisar–India: 59-60.
- Kim KU, Shin DH 2003 The importance of allelopathy in breeding new cultivars. In Weed Management for Developing Countries. *FAO Plant Production and Protection Paper No. 120 (addendum 1)* eds. Labrada R (Rome: Food and Agriculture Org.): 195–210.
- Kruse M, Strandberg M and Strandberg B2000 Ecological Effects of Allelopathic Plants a Review.National Environmental Research Institute, Silkeborg, Denmark.NERI Technical Report No. 315: 66 pp.
- Lin J, Smith Jr. RJ, Dilday RH1992 Comparison of allelopathic rice and bensulfuron for aquatic weed control in rice.

- WSSA Abstr.33: 170.
- Maguire JD1962 Speed of germination—aid in selection and evaluation for seedling emergence and vigor. Crop Sci. 2:176–177
- Olefsdotter M, Navarez D, Moody K1995Allelopathic potential in rice (*Oryza sativa L.*) gremplasm. Ann. Appl. Biol. 127:543–560.
- Reigosa M J, Sanchez-Moreiras A and Gonzalez L 1999 Ecolophysiological approach in allelopathy. Crit. Rev. Plant Sci.18: 577–608.
- Rice EL 1995 Biological control of weeds and plant diseases: advances in applied allelopathy. Norman (Okla., USA): University of Oklahoma Press: 448.
- Rice EL 1984 Allelopathy. 2nd ed. Academic Press, Orlando, FL: 422.
- Valverde BE, Riches CR and Caseley JC 2000Prevention and management of herbicideresistant weeds in rice. Published by Grafos, S.A., Cartago, Costa Rica: 25-30.
- Wathugala DL, Jayakody TSD, Ranagalage AS and Kumarasinghe HMS 2013 Evaluation of allelopathic potential of Sri Lankan improved rice (Oryza sativa L.) varieties for control of *Echinochloa crus-galli.Research on crops* 14(4): 985-994.
- Weir TL, Park SW, Vivanco JM 2004 Biochemical and physiological mechanisms mediated by allelochemicals. Current opinion plant biology J. 9: 195-203.